

# SVENSK STANDARD

## SS-EN ISO 20963:2011



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### **Markundersökningar – Effekter av föroreningar på insektslarver (Oxythyrea funesta) – Bestämning av akut toxicitet (ISO 20963:2005)**

### **Soil quality – Effects of pollutants on insect larvae (Oxythyrea funesta) – Determination of acute toxicity (ISO 20963:2005)**

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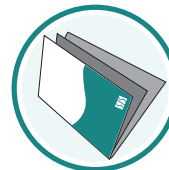
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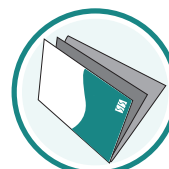
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Denna standard ersätter SS-ISO 20963:2005, utgåva 1.

The European Standard EN ISO 20963:2011 has the status of a Swedish Standard. This document contains the official version of EN ISO 20963:2011.

This standard supersedes the Swedish Standard SS-ISO 20963:2005, edition 1.

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EUROPEAN STANDARD

**EN ISO 20963**

NORME EUROPÉENNE

EUROPÄISCHE NORM

June 2011

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ICS 13.080.30

English Version

## Soil quality - Effects of pollutants on insect larvae (*Oxythyrea funesta*) - Determination of acute toxicity (ISO 20963:2005)

Qualité du sol - Effets des polluants vis-à-vis des larves d'insectes (*Oxythyrea funesta*) - Détermination de la toxicité aiguë (ISO 20963:2005)

Bodenbeschaffenheit - Auswirkungen von Schadstoffen auf Insektenlarven (*Oxythyrea funesta*) - Bestimmung der akuten Toxizität (ISO 20963:2005)

This European Standard was approved by CEN on 10 June 2011.

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This European Standard exists in three official versions (English, French, German). A version in any other language made by translation under the responsibility of a CEN member into its own language and notified to the CEN-CENELEC Management Centre has the same status as the official versions.

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## **Foreword**

The text of ISO 20963:2005 has been prepared by Technical Committee ISO/TC 190 "Soil quality" of the International Organization for Standardization (ISO) and has been taken over as EN ISO 20963:2011 by Technical Committee CEN/TC 345 "Characterization of soils" the secretariat of which is held by NEN.

This European Standard shall be given the status of a national standard, either by publication of an identical text or by endorsement, at the latest by December 2011, and conflicting national standards shall be withdrawn at the latest by December 2011.

Attention is drawn to the possibility that some of the elements of this document may be the subject of patent rights. CEN [and/or CENELEC] shall not be held responsible for identifying any or all such patent rights.

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### **Endorsement notice**

The text of ISO 20963:2005 has been approved by CEN as a EN ISO 20963:2011 without any modification.

## Introduction

This International Standard describes a method for the determination of the acute toxicity of contaminated soils and chemicals to the larvae of *Oxythyrea funesta*, a phytophagous coleopteran (*Scarabaeidae*, *Cetoniinae*) with wide geographic distribution (Europe, North Africa and the Middle East).

*Oxythyrea funesta* has many characteristics which make it suitable for soil quality monitoring or testing effects of chemicals:

- ecological relevance: this type of organism contributes in many ways to soil structure by stimulating soil aeration and drainage;
- the first stages of development, i.e. incubation of eggs, larval cycle and pupation, are underground;
- the larvae of *Oxythyrea funesta* are tolerant to modifications of the test substrate granulometry;
- this species can be bred under controlled conditions.



# Soil quality — Effects of pollutants on insect larvae (*Oxythyrea funesta*) — Determination of acute toxicity

## 1 Scope

This International Standard describes a method for the determination of the effects of contaminated soils and substances on the survival of the larvae of *Oxythyrea funesta*. The larvae are exposed to the pollutants by cuticular and alimentary uptake.

For contaminated soils, the effects on the survival are determined in the test soil and in a control soil. Depending on the objectives of the study, the control and dilution substrates (dilution series of contaminated soil) are either uncontaminated soil comparable to the soil sample to be tested or artificial soil substrate. Effects of substances are assessed using a defined artificial soil substrate.

This International Standard is not applicable to volatile substances, i.e. substances for which Henry's constant or the air/water partition coefficient is greater than 1, or for which the vapour pressure exceeds 0,001 33 Pa at 25 °C.

NOTE This method does not take into account the possible degradation of the substances or pollutants during the test.

## 2 Normative references

The following referenced documents are indispensable for the application of this document. For dated references, only the edition cited applies. For undated references, the latest edition of the referenced document (including any amendments) applies.

ISO 10381-6, *Soil quality — Sampling — Part 6: Guidance on the collection, handling and storage of soil for the assessment of aerobic microbial processes in the laboratory*

ISO 10390, *Soil quality — Determination of pH*

ISO 11268-1, *Soil quality — Effects of pollutants on earthworms (Eisenia fetida) — Part 1: Determination of acute toxicity using artificial soil substrate*

ISO 11269-2:—<sup>1)</sup>, *Soil quality — Determination of the effects of pollutants on soil flora — Part 2: Effects of chemicals on the emergence and growth of higher plants*

## 3 Terms and definitions

For the purposes of this document, the following terms and definitions apply.

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1) To be published. (Revision of ISO 11269-2:1995)

**3.1**  
**lethal concentration 50**  
**LC 50**  
median lethal concentration of test substance or percent dilution of contaminated soil, which kills 50 % of the test organism within the test period

**3.2**  
**lethal concentration  $x$**   
**LC $_x$**   
concentration of the test substance or percent dilution of contaminated soil which kills  $x$  % of the test organism within the test period

NOTE  $x$  is the percentage (10, 20, 25) of this effect.

**3.3**  
**Lowest Observed Effect Concentration**  
**LOEC**  
lowest tested concentration of test substance or contaminated soil at which a statistically significant effect is observed compared with the control

NOTE All test concentrations above the LOEC have a harmful effect equal or greater than those observed at the LOEC.

**3.4**  
**No Observed Effect Concentration**  
**NOEC**  
highest tested concentration of test substance or contaminated soil at which no statistically significant effect is observed compared with the control

NOTE The NOEC is the test concentration immediately below the LOEC.

**3.5**  
**test mixture**  
mixture of the test substance with the test substrate, mixture of contaminated soil with the test substrate or mixture of contaminated soil with an uncontaminated soil comparable to the soil sample to be tested

**3.6**  
**diapause**  
interruption of metabolism during egg, larva, pupa or imago development

## 4 Principle

Larvae of *Cetoniinae* (species *Oxythyrea funesta*) are exposed to a dilution range of contaminated soil or to a range of concentrations of test substance. The mortality of larvae is determined after 10 days. Test mixtures are prepared at the start of the test and are not renewed within the test period.

The results obtained from the test are compared with a control and are used to determine the concentration which causes mortality of 50 % of the larvae (LC 50<sub>10days</sub>).

The test is conducted in two steps:

- a preliminary test to determine appropriate dilution/concentration range in the final test;
- the definitive test to determine the dilutions/concentrations causing between 10 % and 90 % mortality, which yields the test result.

It may also be possible to determine the effects of contaminated soils or substances on the growth of larvae (optional). The increase in mass within the test period allows this criterion to be considered as complementary to mortality, in order to assess the effects of contaminated soils or substances.

## 5 Test environment

Tests shall be performed at a temperature of  $(26 \pm 1)$  °C in complete darkness.

## 6 Reagents

### 6.1 Biological material

The species used in the test is *Oxythyrea funesta* (*Scarabaeidae*, *Cetoniinae*). Third-instar larvae with a fresh mass within the range 100 mg to 200 mg are required to perform the test. The larvae shall be healthy, without any bites or other visible injuries.

**NOTE** Depending on the breeding conditions, described in Annex A, larvae approximately two weeks old are suitable for the test.

Larvae of similar size shall be selected. The difference in mass between the smallest and the largest larva within a single test container shall not exceed 50 mg.

Eliminate the particles of breeding substrate stuck to the integument using, for example, a soft brush before weighing the larvae. It is also possible to leave the animals to move along on slightly moist paper in order to eliminate the breeding substrate stuck to the integument.

Synchronisation of breeding is necessary. An example of breeding technique for *Oxythyrea funesta* is given in Annex A.

### 6.2 Test substrate

The mass of substrate used per glass container (7.1) shall be equivalent to 300 g (dry mass).

The substrate, called artificial soil, shall have the following composition (in accordance with ISO 11268-1):

- sphagnum peat, air-dried, finely ground and with no visible plant remains: 10 % (expressed on a dry mass basis);
- kaolinite clay containing not less than 30 % kaolinite: 20 % (expressed on a dry mass basis);
- industrial quartz sand (dominant fine sand with more than 50 % particle size between 0,05 mm and 0,20 mm): 70 % (expressed on a dry mass basis).

Add pulverised calcium carbonate ( $\text{CaCO}_3$ ), of recognised analytical grade, as necessary to bring the pH of the wetted substrate to  $6,0 \pm 0,5$  (commonly between 0,5 % and 1 % of the mass of the dry ingredients).

Prepare the artificial soil by mixing the dry constituents listed above thoroughly in a large-scale laboratory mixer. The amount of calcium carbonate required can vary, depending on properties of the individual batch of sphagnum peat, and should be determined by weighing subsamples immediately before the test.

Store the mixed artificial soil at room temperature. To determine pH and the maximum water-holding capacity, pre-moisten the dry artificial soil at least two days before starting the test by adding deionized water to obtain half of the required final water content of 50 % of the maximum water-holding capacity.