

# SVENSK STANDARD

## SS-EN ISO 16072:2011

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### **Markundersökningar – Laboratoriemetod för bestämning av mikrobiell markrespiration (ISO 16072:2002)**

### **Soil quality – Laboratory methods for determination of microbial soil respiration (ISO 16072:2002)**

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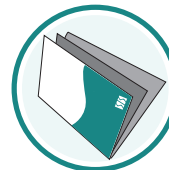
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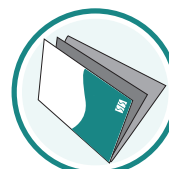
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Denna standard ersätter SS-ISO 16072, utgåva 1.

The European Standard EN ISO 16072:2011 has the status of a Swedish Standard. This document contains the official version of EN ISO 16072:2011.

This standard supersedes the Swedish Standard SS-ISO 16072, edition 1.

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EUROPEAN STANDARD

**EN ISO 16072**

NORME EUROPÉENNE

EUROPÄISCHE NORM

June 2011

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ICS 13.080.30

English Version

## Soil quality - Laboratory methods for determination of microbial soil respiration (ISO 16072:2002)

Qualité du sol - Méthodes de laboratoire pour la détermination de la respiration microbienne du sol (ISO 16072:2002)

Bodenbeschaffenheit - Laborverfahren zur Bestimmung der mikrobiellen Bodenatmung (ISO 16072:2002)

This European Standard was approved by CEN on 10 June 2011.

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<b>Contents</b>	<b>Page</b>
Foreword .....	iv
Introduction .....	v
1 Scope.....	1
2 Normative references .....	1
3 Terms and definitions .....	1
4 Procedure.....	2
4.1 General conditions.....	2
4.2 Choice of the measuring system.....	3
5 Measuring systems .....	3
5.1 Determination of O <sub>2</sub> consumption by static incubation in a pressure-compensation system .....	3
5.2 Determination of CO <sub>2</sub> release by titration in a static system .....	4
5.3 Coulometric determination of CO <sub>2</sub> release in a static system .....	6
5.4 Determination of CO <sub>2</sub> release using an infrared gas analyser in a flow-through system.....	7
5.5 Determination of CO <sub>2</sub> release using gas chromatography in a flow-through system and a static system .....	10
5.6 Determination of soil respiration by pressure measurement in a static system .....	15
Bibliography .....	19

## **Foreword**

The text of ISO 16072:2002 has been prepared by Technical Committee ISO/TC 190 "Soil quality" of the International Organization for Standardization (ISO) and has been taken over as EN ISO 16072:2011 by Technical Committee CEN/TC 345 "Characterization of soils" the secretariat of which is held by NEN.

This European Standard shall be given the status of a national standard, either by publication of an identical text or by endorsement, at the latest by December 2011, and conflicting national standards shall be withdrawn at the latest by December 2011.

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The text of ISO 16072:2002 has been approved by CEN as a EN ISO 16072:2011 without any modification.

## Introduction

This International Standard is derived from the German standard DIN 19737 (see [1]). It describes methods for the determination of microbial soil respiration in the laboratory.

Microbial soil respiration results from the mineralization of organic substances. In this process, organic substances are oxidized to the end products carbon dioxide and water, with concurrent uptake of O<sub>2</sub> for aerobic microorganisms. The soil respiration is measured by the determination of O<sub>2</sub> consumption and/or by CO<sub>2</sub> release. Respiration is a measure of the overall activity of soil microorganisms.



# Soil quality — Laboratory methods for determination of microbial soil respiration

## 1 Scope

This International Standard describes methods for the determination of soil microbial respiration of aerobic, unsaturated soils. The methods are suitable for the determination of O<sub>2</sub> uptake or CO<sub>2</sub> release, either after addition of a substrate (substrate-induced respiration), or without substrate addition (basal respiration).

This International Standard is applicable to the measurement of soil respiration in order to:

- determine the microbial activity in soil (see [3]);
- establish the effect of additives (nutrients, pollutants, soil improvers, etc.) on the metabolic performance of microorganisms;
- determine the microbial biomass (see [4]);
- determine the metabolic quotient  $q\text{CO}_2$ .

## 2 Normative references

The following referenced documents are indispensable for the application of this document. For dated references, only the edition cited applies. For undated references, the latest edition of the referenced document (including any amendments) applies.

ISO 10381-6:1993, *Soil quality — Sampling — Guidance on the collection, handling and storage of soil for the assessment of aerobic microbial processes in the laboratory*

ISO 11274:1998, *Soil quality — Determination of the water-retention characteristic — Laboratory methods*

ISO 11465:1993, *Soil quality — Determination of dry matter and water content on a mass basis — Gravimetric method*

## 3 Terms and definitions

For the purposes of this document, the following terms and definitions apply.

### 3.1

#### **basal respiration**

microbial soil respiration without addition of nutrients

### 3.2

#### **substrate-induced respiration**

#### **SIR**

microbial soil respiration after addition of nutrients

NOTE      Glucose is an example of an added nutrient.

### 3.3

#### microbial activity

metabolic performance of microorganisms

NOTE It can be measured, for example, as O<sub>2</sub> uptake or CO<sub>2</sub> release.

### 3.4

#### metabolic quotient

$q_{CO_2}$

specific metabolic activity of soil microorganisms, which can be calculated as the quotient basal respiration: microbial biomass

NOTE Metabolic quotient is usually expressed as milligrams of CO<sub>2</sub> carbon released per hour per gram of microbial biomass carbon.

### 3.5

#### rate of CO<sub>2</sub> formation [O<sub>2</sub> consumption]

$R_{CO_2}$  [ $R_{O_2}$ ]

amount of CO<sub>2</sub> released [O<sub>2</sub> consumed] per time unit from a mass unit of soil

NOTE 1 Soil respiration is usually measured as the rate of CO<sub>2</sub> formation or O<sub>2</sub> consumption.

NOTE 2 It is usually expressed as milligrams CO<sub>2</sub> [or O<sub>2</sub>] per gram per hour (mg CO<sub>2</sub> [or O<sub>2</sub>]·g<sup>-1</sup>·h<sup>-1</sup>).

### 3.6

#### microbial biomass

mass of intact microbial cells in a given soil

NOTE This is usually estimated from the measurement of carbon or nitrogen content of these cells.

## 4 Procedure

### 4.1 General conditions

#### 4.1.1 Soil sampling and storage

Sample, store and pre-incubate soils in accordance with ISO 10381-6, independently of the choice of the procedure and the respiration parameter to be measured (basal respiration, SIR).

#### 4.1.2 Measuring and incubation conditions

Soil respiration is strongly influenced by water content and temperature. Therefore these parameters should be recorded in the final report. At suction pressures > 0,03 MPa, the soil respiration will decrease considerably. The water content of the test soil is optimal when it corresponds with a pore water pressure of – 0,01 MPa to – 0,03 MPa (measured with an accuracy of 5 %, in accordance with ISO 11274) or 40 % to 60 % of the maximum water-holding capacity, respectively. A stable temperature should be used. Incubation temperatures between 20 °C and 30 °C are generally recommended, but other temperatures may be used if required. In the description of the methods, examples of incubation temperatures are given as well as the accuracy of temperature maintenance and measurement.

If a method is used for the determination of soil microbial biomass, a temperature of 22 °C is recommended because biomass calculations have been calibrated to this temperature.

When soil samples are compared with respect to soil respiration, they should have the same moisture status (pore water pressure or percentage of maximum water-holding capacity).

## 4.2 Choice of the measuring system

Each measurement method has its own advantages and disadvantages. Care is needed, because the results obtained by O<sub>2</sub> uptake and by CO<sub>2</sub> release are not strictly compatible. It is the responsibility of the investigator to decide which of these methods is to be used.

One of the systems described in Clause 5 should be used.

Systems for measuring CO<sub>2</sub> do not distinguish between CO<sub>2</sub> released from microbial activities and CO<sub>2</sub> resulting from abiotic processes. For alkaline soils and soils with a high organic matter content, which can release considerable amounts of abiotically released CO<sub>2</sub>, methods using O<sub>2</sub> uptake are recommended.

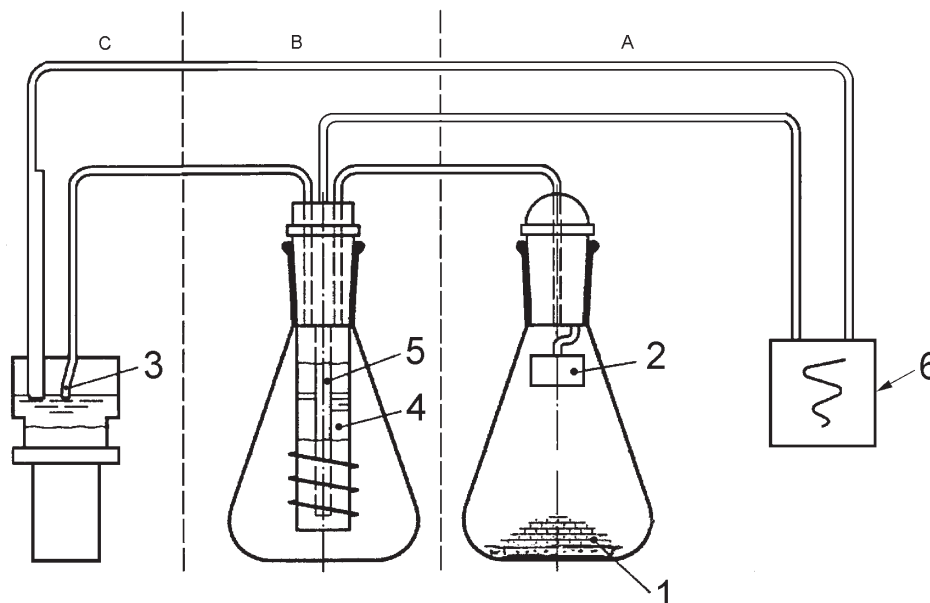
NOTE The advantages and disadvantages are described in the individual descriptions of the methods.

## 5 Measuring systems

### 5.1 Determination of O<sub>2</sub> consumption by static incubation in a pressure-compensation system

#### 5.1.1 Principle

The determination is based on the measurement of O<sub>2</sub> consumption during incubation of a soil sample in a closed system. The O<sub>2</sub> in the system is replenished electrochemically. The CO<sub>2</sub> released is absorbed by calcium hydroxide [Ca(OH)<sub>2</sub>].



#### Key

A	reaction vessel	1	soil sample	4	electrolyte
B	oxygen generator	2	CO <sub>2</sub> absorbent	5	electrodes
C	pressure indicator	3	pressure cell	6	recorder with display

Figure 1 — Determination of O<sub>2</sub> consumption (showing connection of a measuring unit)

#### 5.1.2 Apparatus

A detailed description of the apparatus can be found in [4]; the essential features are as follows.