

# SVENSK STANDARD

## SS-EN 16802:2016



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**Livsmedelsanalyser – Bestämning av grundämnen och deras kemiska species – Bestämning av oorganisk arsenik i livsmedel av marint och vegetabiliskt ursprung med anjonbytes HPLC-ICP-MS**

**Foodstuffs – Determination of elements and their chemical species – Determination of inorganic arsenic in foodstuffs of marine and plant origin by anion-exchange HPLC-ICP-MS**

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EUROPEAN STANDARD

EN 16802

NORME EUROPÉENNE

EUROPÄISCHE NORM

March 2016

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English Version

Foodstuffs - Determination of elements and their chemical species - Determination of inorganic arsenic in foodstuffs of marine and plant origin by anion-exchange HPLC-ICP-MS

Produits alimentaires - Détermination des éléments et de leurs espèces chimiques - Détermination de la teneur en arsenic inorganique dans les produits alimentaires d'origines marine et végétale, par CLHP avec échange d'anions et spectrométrie de masse à plasma induit par haute fréquence (ICP-SM)

Lebensmittel - Bestimmung von Elementen und ihren Verbindungen - Bestimmung von anorganischem Arsen in Lebensmitteln marinen Ursprungs und pflanzlichen Lebensmitteln mit Anionenaustausch-HPLC-ICP-MS

This European Standard was approved by CEN on 8 February 2016.

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## European foreword

This document (EN 16802:2016) has been prepared by Technical Committee CEN/TC 275 “Food analysis - Horizontal methods”, the secretariat of which is held by DIN.

This European Standard shall be given the status of a national standard, either by publication of an identical text or by endorsement, at the latest by September 2016, and conflicting national standards shall be withdrawn at the latest by September 2016.

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## 1 Scope

This European Standard describes a procedure for the determination of inorganic arsenic in foodstuffs of marine and plant origin by anion-exchange HPLC-ICP-MS following waterbath extraction.

This method has been validated in an interlaboratory test on white rice, wholemeal rice, leek, blue mussels, fish muscle and seaweed with an inorganic arsenic mass fraction in the range 0,073 mg/kg to 10,3 mg/kg [1].

## 2 Normative references

The following documents, in whole or in part, are normatively referenced in this document and are indispensable for its application. For dated references, only the edition cited applies. For undated references, the latest edition of the referenced document (including any amendments) applies.

EN 13804, *Foodstuffs — Determination of elements and their chemical species — General considerations and specific requirements*

EN ISO 3696, *Water for analytical laboratory use — Specification and test methods (ISO 3696)*

## 3 Principle

This standard describes a method for the determination of inorganic arsenic. Inorganic arsenic consists of arsenite, As(III) and arsenate, As(V). A representative test portion of the sample is treated with a diluted nitric acid and hydrogen peroxide solution in a heated waterbath. Hereby the arsenic species are extracted into solution and As(III) is oxidized to As(V). The inorganic arsenic is selectively separated from other arsenic compounds using anion exchange HPLC (High Performance Liquid Chromatography) coupled online to the element-specific detector ICP-MS (Inductively Coupled Plasma Mass Spectrometry) for the determination of the mass fraction of inorganic arsenic. External calibration with solvent matrix-matched standards is used for quantification of the amount of inorganic arsenic.

**WARNING — The use of this standard can involve hazardous materials, operations and equipment. This standard does not purport to address all the safety problems associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to its use.**

## 4 Reagents

### 4.1 General

The concentration of the arsenic species in the reagents and water used shall be low enough to not affect the results of the determination. When using a method of high sensitivity like ICP-MS, the control of the blank levels of water, acid and other reagents is very important. Generally ultra-pure water complying with ISO 3696 grade 1 (i.e. electrical conductivity below 0,1  $\mu\text{S}/\text{cm}$  at 25 °C) and acid of high purity, e.g. cleaned by sub-boiling distillation, are recommended. Reagents should be of minimum p.a. quality where possible. Special facilities can be used in order to avoid contamination during the steps of preparation and measurement (e.g. uses of laminar flow benches or comparable clean room facilities).

**4.2 Nitric acid** concentrated, mass fraction  $w(\text{HNO}_3) \geq 65\%$ , mass concentration of approximately  $\rho(\text{HNO}_3) = 1,4 \text{ g/ml}$ .

Use only nitric acid available with high purity or perform a clean-up by a sub-boiling distillation in order to avoid potential contamination.



#### 4.3 Hydrogen peroxide, $w(\text{H}_2\text{O}_2) \geq 30 \%$ .

High purity is essential to avoid potential contamination. Commercially available hydrogen peroxide for analysis should be tested for contamination of arsenic.

#### 4.4 Extraction solution 1, $c(\text{HNO}_3) = 0,1 \text{ mol/l}$ in 3 % (V/V) $\text{H}_2\text{O}_2$ .

Pour 800 ml of  $\text{H}_2\text{O}$  and then 6,5 ml of  $\text{HNO}_3$  (4.2) and thereafter 100 ml of  $\text{H}_2\text{O}_2$  (4.3) into a 1 000 ml volumetric flask. Fill it up to the mark with  $\text{H}_2\text{O}$ . This solution should be prepared on the same day of use.

It is recommended that the total volume needed for the analysis is estimated and only this amount is produced.

#### 4.5 Extraction solution 2, $c(\text{HNO}_3) = 0,2 \text{ mol/l}$ in 6 % (V/V) $\text{H}_2\text{O}_2$ .

Pour 70 ml of  $\text{H}_2\text{O}$ , 1,3 ml of nitric acid (4.2) and 20 ml of hydrogen peroxide (4.3) into a 100 ml volumetric flask. Fill up to the mark at 100 ml with  $\text{H}_2\text{O}$ . This solution should be prepared on the same day of use.

It is recommended that the total volume needed for the analysis is estimated and only this amount is produced.

#### 4.6 Ammonium carbonate, $w[(\text{NH}_4)_2\text{CO}_3] \geq 99,999 \%$ , for production of mobile phase solution.

#### 4.7 Aqueous ammonia, $w[\text{NH}_3(\text{aq.})] \geq 25 \%$ , for adjustment of pH in the mobile phase.

#### 4.8 Methanol, ( $\text{CH}_3\text{OH}$ ), HPLC grade, for production of mobile phase.

#### 4.9 Mobile phase, e.g. 50 mmol/l ammonium carbonate in 3 % methanol at pH 10,3.

Dissolve e.g. 4,80 g of ammonium carbonate (4.6) in approximately 800 ml of water. Adjust the pH to 10,3 with aqueous ammonia (4.7), add 30 ml of methanol (4.8) and fill up to 1 000 ml with water. Filter the mobile phase solution through a 0,45  $\mu\text{m}$  filter prior to use.

The optimal concentration of ammonium carbonate in the mobile phase depends on the analytical column used (e.g. brand, particle size and dimensions). The appropriate concentration of ammonia carbonate is at the discretion of the analyst and should fulfil the criteria for sufficient resolution of the arsenate peak as stated in 5.10.

Methanol is added to the mobile phase in order to enhance the signal intensity for arsenic (carbon enhancement effect [2]). The concentration of methanol for maximum signal depends on the instrument used and should be identified by the analyst.

#### 4.10 Diarsenic trioxide, $w(\text{As}_2\text{O}_3) \geq 99,5 \%$ , optional.

#### 4.11 Potassium hydroxide solution, $\rho(\text{KOH}) = 20 \text{ g/100 ml}$ , optional.

#### 4.12 Sulfuric acid solutions, $w(\text{H}_2\text{SO}_4) = 20 \%$ and $w(\text{H}_2\text{SO}_4) = 1 \%$ , optional.

#### 4.13 Phenolphthalein, optional.

#### 4.14 Standard solutions, with an arsenic mass concentration of 1 000 mg/l.

The use of commercial standards of arsenic, arsenic III and/or V, with a mass concentration of 1 000 mg/l is recommended.

Otherwise proceed as follows: Dissolve e.g. 1,320 g of diarsenic trioxide (4.10) in 25 ml of potassium hydroxide solution (4.11), neutralize with 20 % sulfuric acid solution (4.12) with phenolphthalein (4.13) as indicator and dilute to 1 000 ml in a volumetric flask with 1 % sulfuric acid solution (4.12).

NOTE By preparing the standard in the extraction solution 1 (4.4) all arsenite will be completely oxidized to arsenate.

#### 4.15 Calibration solutions.

Prepare a range of standards including a blank calibration solution that covers the linear range of the analyte to be determined by diluting the analyte stock solution with extraction solution 1 (4.4). Appropriate matrix matching of the calibration solutions shall be performed by using the extraction solution 1 (4.4) for the final dilution step, which furthermore will prevent reduction of arsenate to arsenite. Transfer an aliquot of the calibration solutions to HPLC vials prior to analysis (6.3.2).

The quantitative oxidation of arsenite to arsenate in the standard solutions should be verified (e.g. visual inspection of chromatogram by looking for an additional peak or a reduced intensity of the arsenate peak).

**4.16 Solution for checking chromatographic separation**, containing the organic arsenic compounds (e.g. 10 µg/l) monomethylarsenous acid (MMA), dimethylarsinic acid (DMA) and arsenobetaine (AB), as well as arsenate (e.g. 10 µg/l) and chloride (e.g. 100 mg/l).

## 5 Apparatus and equipment

### 5.1 General

To minimize the contamination, all apparatus and equipment that come into direct contact with the sample and the solutions shall be carefully pre-treated. It is recommended to avoid the use of glassware, since this may cause contamination with arsenate, see [3].

**WARNING** — Some autosampler systems use syringes made of glass. In this case, it is only possible to check for contamination and to minimize it.

**5.2 Laboratory grinder**, capable of grinding to a particle size of less than 0,5 mm.

**5.3 Analytical balance**, accuracy of 1 mg.

**5.4 Filtering device**, for filtration of mobile phase, maximum pore size 0,45 µm.

**5.5 Waterbath**, adjustable to 90 °C.

**5.6 Centrifuge**, minimum 2 010 g (4 000 min<sup>-1</sup>).

**5.7 Single use syringe filters (0,45 µm) or HPLC vials with filters**, compatible with acidic solutions for filtering of test solutions prior to analysis.

**5.8 Plastic volumetric flasks**, for preparation of mobile phase, blank and calibration solutions.

NOTE If calibration standards are prepared by weighing, plasticware without marks can be used.

**5.9 High Performance Liquid Chromatographic System (HPLC).**

**5.10 Strong anion exchange column (SAX)**, suitable for selective separation of arsenate from other arsenic compounds present in the sample extracts.

It is advisable to use a guard column to prolong the life-time of the analytical column.

Usually, the minimum acceptable retention time for the analyte is twice the retention time corresponding to the void volume of the column. Furthermore the nearest peak in the chromatogram should be separated from the analyte peak by at least one full peak width at 10 % of the analyte peak height. It is recommended to verify sufficient separation of the analyte peak using a solution of organic arsenic compounds (e.g. monomethylarsenous acid (MMA), dimethylarsinic acid (DMA) and arsenobetaine (AB)) and arsenate. Make sure that the HPLC run is long enough for chloride ( $m/z$  35) and for arsenic compounds with longer retention times than arsenate, to elute from the column prior to injection of the next sample. It should furthermore be ensured that the arsenate and chloride peaks do not co-elute in order to avoid interference from the polyatomic ion  $^{40}\text{Ar}^{35}\text{Cl}^+$  in the mass spectrometer. Examples of chromatographic SAX columns used in the collaborative trial can be found in Annex B, see also [1].

### **5.11 Inductively coupled plasma mass spectrometer (ICP-MS).**

**5.12 Argon gas**, purity  $\geq 99,99$  %.

## **6 Procedure**

### **6.1 General**

Follow the procedures for sample preparation as given in EN 13804.

### **6.2 Waterbath extraction**

Weigh a test portion of approximately 0,2 g to 0,5 g sample in powder form to the nearest milligram into a tube and fill up to 10,00 ml with extraction solution 1 (4.4). Include also a reagent blank sample. The tubes shall be securely closed with a tight lid. Shake the tubes thoroughly in order to wet the sample sufficiently with the extraction solution 1 (4.4). Some finely powdered samples may need extended wetting time (e.g. overnight).

If a fresh sample is extracted, the water content shall be taken into account. The sample weight should correspond to 0,2 g to 0,5 g dry matter. The concentration of extraction solution should be adjusted accordingly, keeping the matrix matching at the same level. Proceed e.g. as follows: weigh in the test sample, add water up to 5 ml, mix thoroughly, add 5 ml of extraction solution 2 (4.5) and mix again.

The solutions are then placed in a heated waterbath at  $90\text{ }^\circ\text{C} \pm 2\text{ }^\circ\text{C}$  and extracted for  $60\text{ min} \pm 5\text{ min}$ .

Following the waterbath extraction step, let the samples be cooled to room temperature and subsequently centrifuge the tubes ( $10\text{ min}$ ,  $4\ 000\text{ min}^{-1}$  ( $2\ 010\text{ g}$ )). The supernatant, when transferred to a clean container, can usually be stored in a refrigerator (at approximately  $4\text{ }^\circ\text{C}$ ) for a maximum of one week until analysis (6.3.3). Prior to analysis, all sample extracts should be filtered (5.7) into HPLC vials.

### **6.3 Determination of inorganic arsenic by HPLC-ICP-MS**

#### **6.3.1 Preparation of HPLC-ICP-MS for analysis**

The HPLC and ICP-MS operating conditions shall be based on the general information provided by the manufacturer of the instruments taking into account the operating conditions of the analytical column.

An arsenic solution, e.g.  $10\text{ }\mu\text{g/l}$  arsenate in 3 % methanol, may be used to optimize the test system according to the manufacturer's instructions. Arsenic is mono-isotopic and can be evaluated at a mass/charge ratio ( $m/z$ ) of 75.

It is advisable to allow the HPLC system (incl. the analytical column) to equilibrate and ensure stable conditions by turning on the HPLC mobile phase flow in advance prior to start of the analysis. Repeated injections of a sample extract may be necessary until stable chromatography is achieved and the